HPLC Method Development Guidelines

REVERSED PHASE HPLC METHOD DEVELOPMENT PARAMETERS

- Analyte structure and pK
- Mobile phase composition
- HPLC column choice (optimal stationary phase)

RETENTION MECHANISMS

Hydrophobic / Non-polar / Reversed Phase **Typical Phases:** C18, C8, Phenyl, Polymer (i.e. SDVB) Interactions with alkyl chains



Van der Waals Forces

Ion-Exchange

Typical Phases: SCX, WCX, SAX, NH, Interactions with ionic functional groups and strong/weak ion-exchangers



Electrostatic and Ionic Interactions

Hydrophilic / Polar / Normal Phase

Typical Phases: Silica, NH_a, CN Interactions with silanols and polar functional groups



Hydrogen bonding and dipole-dipole interactions

Bond Energies Involved with Retention Mechanisms

Physical Interaction	Types of Compounds	Energy (kJ/mol)
Van der Waals Forces	Alkyl chains, aromatic rings	2 – 10
Dipole-Dipole Interactions	Carbonyls, heterocyclic	8 – 15
Hydrogen Bond	Alcohol, amine, acid	20 – 50
Coulomb Forces	lonic compounds	100 – 400
Covalent Bond	Nonmetals	> 300

TIP:

For aromatic compounds, phenyl phases offer a different selectivity then alkyl phases (C18, C8). Based on the interactions between the pi-electrons of the stationary phase's phenyl ring and the double bonds in the aromatic ring(s) of your analyte, retention time and elution order can change significantly and thus have a very positive effect on your method.

You can further adjust the selectivity of a phenyl stationary phase by using methanol instead of acetonitrile within the mobile phase. The pi-electrons of acetonitrile can interact with both the phenyl phase and aromatic functional groups in a competing way, thus leading to a reduced analyte retention time.



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more buffer details.

Buffer, pK_S and pH Calculated according to the Henderson-Hasselbalch dissociation constant (K_a)

K

р

Buffer* Trifluoroacet Phosphoric A Citric Acid (pk **Formic Acid** Citrate (pK) **Acetic Acid** Citrate (pK) Carbonate (Phosphate (Triethanolam TRIS Diethanolam Ammonia Ethanolamin Carbonate (Diethylamine

Triethvlamin

Piperidin

Practical HPLC Method Development; L.R. Snyder, JJ Kirkland, and JL Glajch. Wiley Interscience, 1997. Introduction to Protein and Peptide HPLC; TP Bradshaw, Phenomenex, 1998. * Common buffers are in bold

Miscibi



MOBILE PHASE COMPOSITION: BUFFER AND ORGANIC SOLVENTS

Henderson-Hasselbalch

By using a pH value which is at least two units above or below the pK value of analyte you ensure that your compound is either fully charged or uncharged.

Reminder: A buffer always consists of an acid or base and the corresponding salt. Typical concentrations are usually in the range of 10-50 mM depending on the needed ionic strength and solubility of the buffer system. See below for

-	[H⁺][A⁻]	
a	[HA]	
LJ _	nK log -	[HA]
	$pn_a - log$	[A ⁻]

[A ⁻]/[HA]	рН
100:1	$pK_a + 2$
10:1	р <i>К</i> _а + 1
1:1	р <i>К</i> _а
1:10	р <i>К</i> _а - 1
1:100	р <i>К</i> _а - 2

Buffer Selection

	Buffer Range	MS
р <i>К</i> а	(pH)	Compatibility
< 2	< 2.5	• ^{**}
2.1	1.5 - 2.7	
3.1	2.5 - 3.7	
3.8	2.8 - 4.8	•
4.7	4.1 - 5.3	
4.8	3.8 - 5.8	•
5.4	4.8 - 6.0	
6.4	5.4 - 7.4	•
7.2	6.6 - 7.8	
7.8	6.8 - 8.8	•
8.3	7.3 - 9.3	
8.9	7.9 - 9.9	•
9.2	8.2 - 10.2	•
9.5	8.5 - 10.5	•
10.3	9.3 - 11.3	•
10.5	9.5 - 11.5	•
11.0	10.0 - 12.0	•
11.1	10.1 - 12.1	
	р <i>К</i> а < 2 2.1 3.1 3.8 4.7 4.8 5.4 6.4 7.2 7.8 8.3 8.9 9.2 9.5 10.3 10.5 11.0 11.1	Buffer Range pKa (pH) < 2

** Low concentrations of TFA may be used in LC/MS applications, however it can affect sensitivity.

Solvent Strength



TIP:

Use both methanol and acetonitrile to vary the selectivity.

lity T	abl	le															Solvent	Polarity Index	Refractive Index @ 20 °C	UV (nm) Cutoff @ 1 AU	Boiling Point (°C)	Viscosity (cPoise)	Solubility in Water (% w/w)		
																	Acetic Acid	6.2	1,372	230	118	1.26	100		
																	Acetone	5.1	1,359	330	56	0.32	100		
																	Acetonitrile	5.8	1,344	190	82	0.37	100		
																	Benzene	2.7	1,501	280	80	0.65	0.18		
																	Butyl Acetate	4.0	1,394	254	125	0.73	0.43		
																	n-Butanol	3.9	1,399	215	118	2.98	7.81		
																	Carbon tetrachloride	1.6	1,466	263	77	0.97	0.08		
																	Chloroform	4.1	1,446	245	61	0.57	0.815		
																	Cyclohexane	0.2	1,426	200	81	1.00	0.01		
																	1,2-Dichloroethane ¹	3.5	1,444	225	84	0.79	0.81		
																	Dichloromethane ²	3.1	1,424	235	41	0.44	1.6		
																	Dimethylformamide	6.4	1,431	268	155	0.92	100		
																	Dimethyl Solfoxide ³	7.2	1,478	268	189	2.00	100		
																	Dioxane	4.8	1,422	215	101	1.54	100		
																	Ethyl Acetate	4.4	1,372	260	77	0.45	8.7		
																	Ethanol	5.2	1,360	210	78	1.20	100		
																	di-Ethyl Ether	2.8	1,353	220	35	0.32	6.89		
																	Heptane	0.0	1,387	200	98	0.39	0.0003		
																	Hexane	0.0	1,375	200	69	0.33	0.001		
																	Methanol	5.1	1,329	205	65	0.60	100		
																	Methyl-t-Butyl Ether ⁴	2.5	1,369	210	55	0.27	4.8		
																	Methyl Ethyl Ketone ^₅	4,7	1,379	329	80	0.45	24		
																	Pentane	0.0	1,358	200	36	0.23	0.004		
																	n-Propanol	4.0	1,384	210	97	2.27	100		
																	iso-Propanol ⁶	3.9	1,377	210	82	2.30	100		
																	di-iso-Propyl Ether	2.2	1,368	220	68	0.37			
																	Tetrahydrofuran	4.0	1,407	215	65	0.55	100		
																	Toluene	2.4	1,496	285	111	0.59	0.051		
																	Trichloroethylene	1.0	1,477	273	87	0.57	0.11		
																	Water	9.0	1,333	200	100	1.00	100		
																	Xylene	2.5	1,500	290	139	0.61	0.018		
er				Je ³	ide	ne ¹		ride						& Misc	ible		SYNONYM TAE	BLE de ⁴ ter	t-Butvl Methyl Ether						
Propyl Eth Ipanol ⁶ anol e	Ethyl Ketu -t-Butyl Et	lor		l Ether	Cetate	- e	nyl Sulfoxia	nylformam	thereafter the the	exane	form tetrachlo	nol	le	itrile	e.	Acid	Immiscible [*]	ŧ			² Methylene Chlo ³ Methyl Sulfoxic	oride ⁵ 2-1 le ⁶ 2-1	⁵ 2-Butanone ⁶ 2-Propanol		
dı-ıso- iso-pro n-Prop Pentan	Methyl	Methar	Hexan(di-Ethy	Ethano	Dioxan	Dimeth	Dimet	1.2-Dic	Cycloh	Chlorot Carbon	n-Buta	Benzer	Aceton	Aceton	Acetic.	*Immiscible means tha	t in some prop	portions two phases	will be produced.					

ACHIEVE THE BEST RESOLUTION with proper selectivity

Kinetex[®] Core-Shell Phases





Used under HILIC running conditions, this phase provides the highest polar selectivity or retention and separation of hydrophilic compounds

compounds

Aeris[™] Core-Shell Phases WIDEPORE for proteins larger than 10 kDa



PERFORMANCE GAINS ON ANY LC SYSTEM





Kinetex C18





Kinetex Phenyl-Hexyl hvdrocarbon



Kinetex PFP The electronegative fluorine groups offer high selectivity for cationic compounds. Columns are pH stable from 1.5-10 under isocratic conditions. Columns are pH stable 1.5-8.5 under gradient conditions.

Use these different selectivities for the analysis of small compounds up to 10 kDa







Balanced C18 phase that provides the highest degree of hydrophobic selectivity relative to the other Kinetex phases

Aromatic and moderate hydro-

phobic selectivity result in the

of aromatic hydrocarbons

great retention and separation



Pentafluorophenyl phase offers a high degree of steric interactions for improved separation of structural isomers, and the electronegative fluorine groups can offer increased retention o polar basic compounds



Moderate hydrophobic and steric selectivity is offered, bringing ultrahigh performance to USP L7 and other octyl silane methods

Use the Aeris PEPTIDE for the peptides and proteins up to 10 kDa and use the Aeris

• Aeris PEPTIDE and WIDEPORE



- Aeris WIDEPORE
- Large proteins
- Moderately hydrophobic proteins Monoclonal antibodies
- High temperature separations



- Aeris WIDEPORE
- Very large proteins
- Very hydrophobic proteins
- Membrane proteins
- Least retentive



.1 mm ID Kinetex columns are pressure stable up to 1000 bar. When using Kinetex 1.3 µm or 1.7 µm, increased performance can be achieved, however high pressure-capable instrumentation is required.

