

APPLICATIONS

Rapid, Automated Extraction and LC/MS/MS Analysis of Tricyclic Antidepressants from Plasma using Strata™ -X-Drug B SPE and a Kinetex® Core-Shell HPLC/UHPLC Column

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Using an automated SPE method that requires no condition or equilibration steps, time and solvent are saved during the analysis of 9 tricyclic antidepressant (TCA) compounds from human plasma. The rapid automated procedure proved to be reproducible and provided absolute recoveries >75 % for all TCA compounds studied. Coupled to a rapid LC/MS/MS method using a Kinetex core-shell HPLC/UHPLC column, this method can instantly provide time and solvent savings to high-throughput laboratories.

Introduction

Introduced in 1950, tricyclic antidepressants (TCAs) quickly grew to become a popular solution for treatment of clinical depression. Although many TCAs have been replaced by selective serotonin reuptake inhibitors (SSRIs) and serotonin-norepinephrine reuptake inhibitors (SNRIs), TCAs are still prescribed to treat various types of depression as well as chronic pain.

As the name implies, TCAs are heterocyclic chemical compounds consisting of 3 ring structures and a variety of carbon chain functional groups. TCAs are basic in nature and typically have a pK_a of 8 or higher. A basic pK_a combined with a hydrophobic ring structure makes these compounds excellent targets for extraction via a strong cation-exchange SPE sorbent. After extraction, TCAs can be separated using a reversed phase C18 HPLC column, relying on their functional groups to provide separation and resolution of each compound.

Materials and Methods

Sample Pretreatment

500 μ L of plasma was first diluted with 1 mL of 100 mM Sodium acetate buffer (pH 5.0) spiked with TCAs. The diluted sample was then subjected to an automated SPE protocol (below) which was run on a PerkinElmer MultiPROBE® II.

Solid Phase Extraction:

96-Well Plate: Strata-X-Drug B 30 mg/well

Part No.: 8E-S128-TGB

Condition: NOT REQUIRED

Equilibrate: NOT REQUIRED

Load: 500 μ L pretreated plasma

Wash 1: 0.8 mL 100 mM Sodium acetate (pH 5.0)

Wash 2: 0.8 mL Methanol

Dry: 8 to 10 minutes under maximum vacuum

Elute: 0.4 mL of Methanol/Acetonitrile (50:50) plus 2 % Ammonium hydroxide

Blow Down: To dryness under a slow stream of nitrogen @ 45 °C

Reconstitute: 500 μ L of initial mobile phase

HPLC Conditions

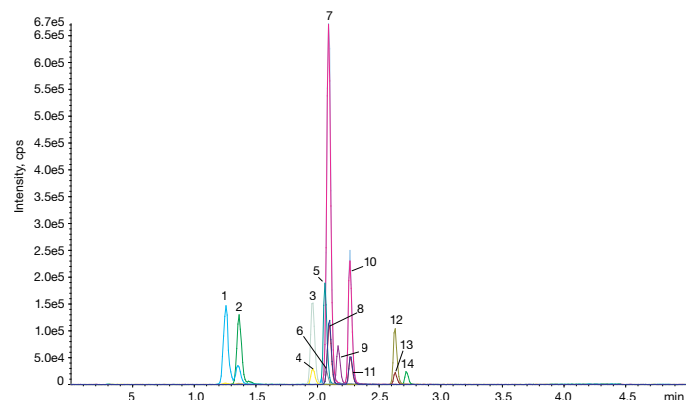
Column: Kinetex 2.6 μ m C18, 100A
Dimensions: 50 x 2.1 mm
Part No.: 00B-4462-AN
Mobile Phase: A: 0.1 % Formic acid in Water
 B: 0.1 % Formic acid in Methanol

Gradient	Time (min)	% B
	0	40
	3.5	80
	4	80
	4.01	40
	5	40

Flow Rate: 500 μ L/min
Temperature: Ambient
Detection: API 4000 MS/MS, ESI Positive (ESI+)
Injection: 10 μ L
Sample:

- Doxepine
- DM-Doxepine
- Imipramine-D3 (IS)
- Imipramine
- Desipramine-D3 (IS)
- Desipramine
- Nortriptyline-D3 (IS)
- Nortriptyline
- Amitriptyline
- Protriptyline-D3 (IS)
- Protriptyline
- Clomipramine-D3 (IS)
- Clomipramine
- DM-Clomipramine

Figure 1. LC/MS/MS chromatogram of 9 TCA compounds using a Kinetex 2.6 μ m C18 core-shell HPLC/UHPLC column



App ID 20937



MS/MS Conditions

Analysis is performed with an Agilent® 1200 HPLC system (Agilent Technologies, Inc., Santa Clara, CA USA) coupled to an AB SCIEX API 4000™ triple-quadrupole tandem mass spectrometer equipped with an ESI probe operating in positive polarity mode. Under an MRM mode, two channels were monitored for 9 TCAs (**Table 1**).

Table 1.
MRM Transitions

Peak Name	MRM Channel
Protriptyline	164.2 ⇒ 191.1
Nortriptyline	264.3 ⇒ 191.2
DM-Doxepin	266.2 ⇒ 107.1
Desipramine	267.2 ⇒ 208.2
Protriptyline-D3	267.2 ⇒ 191.1
Nortriptyline-D3	267.3 ⇒ 191.2
Desipramine-D3	270.2 ⇒ 208.2
Amitriptyline	278.2 ⇒ 191.2
Doxepin	280.3 ⇒ 107.1
Imipramine	281.3 ⇒ 208.2
Imipramine-D3	284.2 ⇒ 208.1
DM-Clomipramine	301.2 ⇒ 242.2
Clomipramine	315.2 ⇒ 242.2
Clomipramine-D3	381.2 ⇒ 242.2

Results and Discussion

The goal of this study was to determine a rapid analysis for 9 TCAs from human plasma that was both sensitive and accurate. When working with human plasma, many endogenous interferences are present including proteins and phospholipids. In order to obtain a clean sample prior to LC/MS/MS analysis, the plasma sample was subjected to an SPE cleanup using Strata™-X-Drug B. Strata-X-Drug B was chosen as the ideal SPE sorbent for our plasma cleanup because it does not require a condition or equilibration step which saved both time and solvent. The sorbent also allowed us to develop a single, simplified method, rather than several different methods for the extraction of 9 TCAs, resulting in absolute recoveries of >75 % for all analytes of interest even at low concentration levels (**Table 2**).

After cleanup by SPE, the TCAs were analyzed by LC/MS/MS using a Kinetex® 2.6 µm C18 core-shell HPLC/UHPLC column. The Kinetex core-shell particles provided the performance, efficiencies and speed of a sub-2 µm column without the added backpressure that is associated with sub-2 µm particles, resulting in a run time of under 3 minutes for all 9 target compounds (**Figure 1**).

Table 2.

Absolute recoveries (%) of TCA compounds after extraction with Strata-X-Drug B SPE




Analyte	500 ng/mL		25 ng/mL	
	Absolute Recovery (%)	% CV	Absolute Recovery (%)	% CV
Amitriptyline	94	5.6	82	12.2
Nortriptyline	80	1.7	75	6.1
Protriptyline	87	2.1	94	4.3
Doxepin	94	7.3	93	3.0
DM-Doxepine	85	4.0	96	9.0
Imipramine	85	1.5	90	4.1
Desipramine	87	1.0	85	1.1
Clomipramine	87	3.4	92	0.2
DM-Clomipramine	84	4.2	89	0.4

Conclusion

Utilizing a single SPE method, 9 TCAs were extracted from plasma and analyzed by LC/MS/MS. The Strata-X-Drug B SPE sorbent did not require a condition or equilibration step which saved both time and solvent. The ability to extract all 9 TCA compounds at once also provided time savings. The extraction resulted in absolute recoveries of >75 % for all 9 analytes at low detection levels (25 ng/mL). Following the extraction, the TCAs were separated and further analyzed using a Kinetex 2.6 µm C18 core-shell HPLC/UHPLC column which provided excellent separation and a fast run time of < 3 minutes. Due to the time and solvent savings, this method could be beneficial in a high-throughput laboratory.

Ordering Information

Strata™-X-Drug B SPE

Format	Sorbent Mass	Part Number	Unit
Tube			
	10 mg	8B-S128-AAK	1 mL (100/box)
	10 mg	8L-S128-AAK†	1 mL (100/box)
	30 mg	8B-S128-TAK	1 mL (100/box)
	30 mg	8L-S128-TAK†	1 mL (100/box)
	30 mg	8B-S128-TBJ	3 mL (50/box)
	60 mg	8B-S128-UBJ	3 mL (50/box)
	60 mg	8B-S128-UCH	6 mL (30/box)
	60 mg	8B-S128-UCL	6 mL (200/box)
Giga™ Tube			
	100 mg	8B-S128-EDG	12 mL (20/box)
96-Well Plate			
	10 mg	8E-S128-AGB	2 Plates/box
	30 mg	8E-S128-TGB	2 Plates/box
	60 mg	8E-S128-UGB	2 Plates/box

†Tab-less tube

Ordering Information

Kinetex® Core-Shell HPLC/UHPLC Columns

5 µm Columns (mm)		SecurityGuard™ ULTRA Cartridges†					SecurityGuard ULTRA Cartridges†
Phases	50 x 2.1	3/pk	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	3/pk
C18	00B-4601-AN	AJO-8782 for 2.1 mm ID	00B-4601-E0	00D-4601-E0	00F-4601-E0	00G-4601-E0	AJO-8768 for 4.6 mm ID

2.6 µm Analytical Columns (mm)						SecurityGuard ULTRA Cartridges†
Phases	30 x 4.6	50 x 4.6	75 x 4.6	100 x 4.6	150 x 4.6	3/pk
C18	00A-4462-E0	00B-4462-E0	00C-4462-E0	00D-4462-E0	00F-4462-E0	AJO-8768 for 4.6 mm ID

2.6 µm MidBore™ Columns (mm)						SecurityGuard ULTRA Cartridges†
Phases	30 x 3.0	50 x 3.0	75 x 3.0	100 x 3.0	150 x 3.0	3/pk
XB-C18	00A-4462-Y0	00B-4462-Y0	00C-4462-Y0	00D-4462-Y0	00F-4462-Y0	AJO-8775 for 3.0 mm ID

2.6 µm Minibore Columns (mm)					SecurityGuard ULTRA Cartridges†
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
C18	00A-4462-AN	00B-4462-AN	00D-4462-AN	00F-4462-AN	AJO-8782 for 2.1 mm ID

1.7 µm MidBore Columns (mm)			SecurityGuard ULTRA Cartridges†
Phases	50 x 3.0	100 x 3.0	3/pk
C18	00B-4475-Y0	00D-4475-Y0	AJO-8775 for 3.0 mm ID

1.7 µm Minibore Columns (mm)					SecurityGuard ULTRA Cartridges†
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
C18	00A-4475-AN	00B-4475-AN	00D-4475-AN	00F-4475-AN	AJO-8782 for 2.1 mm ID

1.3 µm Columns (mm)	
Phase	50 x 2.1
C18	00B-4515-AN

†SecurityGuard ULTRA cartridges require holder, Part No.: AJO-9000



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APPLICATIONS

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Strata-X is patented by Phenomenex. U.S. Patent No. 7,119,145

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